Available online through



www.jbsoweb.com ISSN 2321 - 6328

Research Article

ANTIBACTERIAL ACTIVITY OF FLOWER HEADS OF WEDELIA TRILOBATA (L.) A. S. HITCHC

Rajalakshmi Shankar and Toji Thomas*

Post Graduate and Research Department of Botany, St. Thomas College Palai, Arunapuram, Kerala, India

*Corresponding Author Email: tojidr@yahoo.com

Article Received on: 28/10/14 Accepted on: 25/11/14

DOI: 10.7897/2321-6328.02692

ABSTRACT

Present study is conducted to analyse the antibacterial activity of flower heads of Wedelia trilobata against pathogenic bacterial strains and to analyse phytochemical background of various extracts. Fresh flower heads collected from Kerala, India. Air dried flower heads were extracted successively in petroleum ether, chloroform, acetone, ethanol and water. Preliminary antibacterial activity was analysed by disc-diffusion method and further confirmed by MIC and MBC. Preliminary detection of phytochemicals was done. Among the ten bacterial strains nine showed considerable inhibition of growth towards acetone extract selected for detailed antibacterial evaluation tests like minimum inhibitory concentration (MIC) and minimum bacterial concentration (MBC). The MIC and MBC of floral extracts tested against one of the most sensitive strains, *Streptococcus haemolyticus*. MIC and MBC values were 25 mg/ml and 50 mg/ml respectively. Preliminary phytochemical evaluation revealed the presence of flavonoids, phenolics and terpenoids in active acetone extract. The present investigation showed the effectiveness of crude extract of this plant against tested bacterial strains. The presence of potential phytochemicals like phenolics, flavonoids and terpenoids in active acetone extract might be one of the reasons for its antibacterial property.

Keywords: Wedelia trilobata; antibacterial activity; disc diffusion; phytochemicals.

INTRODUCTION

Wedelia trilobata (L.) A.S. Hitchc, belongs to the family Asteraceae, its common name is Singapore Daisy, synonym is Silphium trilobatum L. The plant is herb, introduced as garden plant, now runs wild and observed as weed. Plant is distributed in all districts of Kerala, India. The plant is a native of Tropical America¹. Antioxidant and antibacterial activities of methanolic extract of flowers of *W. trilobata* were reported². Essential oils isolated from W. trilobata showed antioxidant activity³. Ethanolic extract of *W. trilobata* leaves exhibited antioxidant activity, also indicated its potential in wound healing^{4,5}. Bioactive compounds (ent-kaura-9(11), 16-dien-19-oic acid) from ethanolic extracts of W. trilobata leaves that could influence wound healing⁶. Anthelmintic activity of the various extracts obtained from the leaves of W. trilobata showed significant and dose dependant activity compared to standard drug albendazole, amongst them petroleum ether extract showed better activity⁷. Crude extracts from *W. trilobata* showed antibacterial activity against gram-positive and gram-negative bacteria. None of the tested extracts showed biological activity against fungi⁸. Wedelolides A and B: Novel Sesquiterpene &-Lactones, (9R)-Eudesman-9, 12-olides, from isolated from W. trilobata9. Aqueous extracts of root, stem, leaf and the whole plant of W. *trilobata* showed allopathic potential on rice¹⁰. The present investigation was carried out to analyse antibacterial activity and phytochemical evaluation of flower heads of *W. trilobata*.

MATERIALS AND METHODS Collection of Plant Materials

Fresh specimens (flower heads) of *Wedelia trilobata* (L.) A. S. Hitchc, (Asteraceae) collected from Pala of Kottayam district Kerala, India in month of December 2013. The plant was authenticated by Department of Botany, St. Thomas College, Palai, Kerala, India, where a voucher specimen (RS- 5876) was deposited.

Preparation of Extract

Air dried flower heads were powdered well and used for preparing extracts. 50 g of powdered material extracted in petroleum ether, chloroform, acetone, ethanol and water in the gradation of increasing polarity by keeping the powder in each solvent for 24 hours¹¹. The extraction provided yield of 0.5 %, 1.5 %, 2.5 %, 2.3 % and 1.2 % in petroleum ether, chloroform, acetone, ethanol and water respectively.

Bacterial Strains

Test organisms include Vibrio parahaemolyticus, Salmonella typhi, Bacillus cereus, Salmonella paratyphi, Streptococcus haemolyticas, Proteus vulgaris, Serratia marcescens, Proteus rettgeri, Staphylococcus aureus and Escherichia coli. The species that were not purchased, but originally isolated from clinical materials collected from patients and identified using standard biochemical tests. The bacterial strains were maintained on nutrient agar slants at 4°C in the refrigerator to maintain the stock culture.

Antibacterial Test (Disc-diffusion Method)

The disc-diffusion method as described by Bauer et al.¹² was used to determine the growth inhibition of bacteria in plant extract. Sterile liquid Muller Hinton agar media was transferred into sterile petridish and after solidification; the bacteria were swabbed with a sterile swab under aseptic conditions. Sterile discs prepared using Whatman no. 1 filter paper, of 5mm diameter were used in the study. Test materials were dissolved in the respective solvent to obtain a stock solution of concentration of 500 mg/ml. 10 µL of the solution was loaded per disc to attain a concentration of 5 mg/disc. The discs (including control) were used after drying them in an incubator at 50°C to remove any trace of solvent. Discs were introduced to the medium. The plates were incubated overnight at appropriate incubation temperatures. Microbial growth was conducted in more than three replicates and average inhibitory zone diameter along with standard deviation was recorded. The diameter of the inhibitory zone around each disc was measured. Also sterile disc impregnated with solvents alone were used as control. For each organism average inhibitory zone diameter was determined.

Confirmation of Antibacterial Activity of Active Extract

Antibacterial activity of active extracts was confirmed by determining Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal concentration (MBC). The solidified extract was dissolved in DMSO to make sample concentration of 100 mg/ml

Minimum Inhibitory Concentration (MIC)

The MIC of the extracts was performed by incorporating various amounts (100–0.78 mg/ml) of the extract into sets of test tubes with the culture media. 50 μ L of the bacterial broth culture was added into each of the test tubes. The bacterial cultures containing the plant extracts were incubated at 37°C for 24 hours. Test tube containing only the growth medium and each of the organisms was also incubated under the same conditions as positive controls. The minimum inhibitory concentration was expressed as the lowest concentration of the extracts that did not permit any visible growth when compared to that of the control tubes¹³.

Minimum Bactericidal Concentration (MBC)

Samples from the tubes in previous studies, which did not show any visible growth after a period of incubation, were sub cultured onto a freshly prepared nutrient medium. The minimum bactericidal concentration was taken as the lowest concentration of the extract that did not yield a single colony¹⁴.

Preliminary Phytochemical Analysis of the Extract

The crude samples were subjected to preliminary phytochemical screening for the presence of alkaloids, phenolics, Terpenoids, flavonoids using the method of Harborne¹⁵.

RESULTS

Antibacterial activity and phytochemichal analysis of *Wedelia trilobata* (L.) A.S. Hitchc were conducted using

petroleum ether, chloroform, acetone, ethanol and water as extracting solvents in the gradation of increasing polarity. Out of ten bacterial strains tested, Serratia marcescens did not show any kind of susceptibility to any of extracts. Nine pathogenic strains showed sensitivity towards petroleum ether and acetone extracts (Table 1). Chloroform extract showed antimicrobial activity towards four pathogenic strains such as Vibrio parahaemolyticus, Streptococcus haemolyticus, Escherichia coli and Staphylococcus aureus. Maximum antibacterial activity observed in acetone extract. The extract was highly effective against Streptococcus haemolyticus. Ethanol extract showed antibacterial activity towards five bacterial strains such as Vibrio parahaemolyticus, Streptococcus haemolyticus, Proteus rettgeri, Proteus vulgaris and Staphylococcus aureus. Streptococcus haemolyticus and Staphylococcus aureus showed susceptibility towards water extract of flower heads of the plant. While all other bacteria showed resistance towards various extracts. The Acetone extract was selected for confirmatory antibacterial evaluation tests like Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal (MBC) Streptococcus Concentration towards haemolyticus. The observed MIC and MBC values towards Streptococcus haemolyticus were 25 mg/ml and 50 mg/ml respectively. Preliminary phytochemical evaluation revealed the presence of alkaloids, flavonoids, phenolics and terpenoids. Alkaloids were detected in chloroform and ethanol extracts (Table 2). Flavonoids were found in petroleum ether, chloroform, acetone and ethanol extracts; while water extract was free of alkaloids, flavonoids and terpenoids. Terpenoids were detected in chloroform and acetone extracts.

DISCUSSION

In this work, antibacterial activity of flower head extract of W. trilobata demonstrated against a wide range of bacterial strains. The highest antibacterial activity is being observed against Streptococcus haemolyticus. In recent years multi drug resistant bacteria make therapeutic problems. Plant based antimicrobials have enormous therapeutic potential as they can serve the purpose without any side effects. But side effects are often associated with synthetic antimicrobial compounds. Pharmaceutical companies are now looking for other natural alternative¹⁶. Plants are important source for the development of potential new chemotherapeutic drugs. The broad spectrum of antibacterial activities of the plant extract, possibly due to the identified alkaloids, flavonoids, terpenoids etc. and confirm its use as a health remedy in folklore medicine¹⁷. The antibacterial activity of various plant extracts may be due to the presence of various active principles like alkaloids, terpenoids etc. The bioactivity of plant extracts is attributed to phytochemical constituents for example, alkaloids isolated and characterized from Alstonia rupestris with cytotoxic, antibacterial and antifungal activities¹⁸. Alkaloids isolated from plant are commonly found to have antibacterial and antioxidant properties¹⁹. Terpenoids isolated from plants have antibacterial properties²⁰. Alkaloids, phenols, and flavonoids were observed in various extracts of flower heads of W. trilobata.

	1					
	Zone of inhibition diameter (mm) Extracts used (Concentration - 5 mg/disc)					
Bacterial strains	Petroleum ether	Chloroform	Acetone	Ethanol	Water	
Vibrio parahaemolyticus	11 ± 0.2	10 ± 0.26	14 ± 0.04	13 ± 0.53		
Salmonella paratyphi	10 ± 0.32		13 ± 0.71			
Bacillus cereus	9 ± 0.5		10 ± 0.04			
Salmonella typhi	10 ± 0.05		11 ± 0.20			
Streptococcus haemolyticus	20 ± 0.42	26 ± 0.08	24 ± 0.41	22 ± 0.05	22 ± 0.28	
Proteus vulgaris	9 ± 0.22		13 ± 0.13	10 ± 0.07		
Serratia marcescens						
Proteus rettgeri	13 ± 0.12		14 ± 0.49	8 ± 0.63		
Staphylococcus aureus	11 ± 0.57	17 ± 0.31	16 ± 0.27	9 ± 0.09	8 ± 0.06	
Escherichia coli	13 ± 0.08	9 ± 0.22	12 ± 0.09			

Table 1: Antibacterial activity of flower heads of Wedelia trilobata towards various bacterial strains

Values : Mean \pm Standard Deviation (n = 4)

Table 2: Phytochemichal constituents of flower heads of Wedelia trilobata

Extract used	Alkaloids	Phenolics	Flavonoids	Terpenoids
Petroleum ether	-	+	+	-
Chloroform	+	-	+	+
Acetone	-	+	+	+
Ethanol	+	+	+	-
Water	-	+	-	-

Values : + present, - absent

Active acetone extract showed the presence of phenolics, flavonoids and terpenoids. The antibacterial activity of the active acetone extract of the plant might be due to the presence of these phytochemicals. The present result of antibacterial activity of flower heads of the plant as herbal medicine supported the earlier investigation of antimicrobial activity of the plant reported by Taddeil and Rosas-Romero⁷. Similar result of antibacterial activity was reported from the plant extract by Govindappa *et al*²¹. The present study revealed that acetone extract possessed wide range of antibacterial activity towards gram positive and gram negative bacteria. Therefore acetone extract can be explored for isolating and characterising antibacterial principles involved in antibacterial activity.

CONCLUSION

Flower heads of *Wedelia trilobata* was evaluated for its antibacterial threshold and phytochemical contents in various extracts prepared in the gradation of increase in polarity towards pathogenic bacteria. The flower heads of the plant exhibited antibacterial activity in acetone extract. The acetone extract of the plant showed maximum level of activity towards *Streptococcus haemolyticus*. *Streptococcus haemolyticus* showed maximum level of susceptibility towards various extracts. The presence of phenolics, flavonoids and terpenoids in active acetone extract; acetone extract exhibited minimum inhibitory concentration as 25 mg/ml and minimum bactericidal concentration as 50 mg/ml towards *Streptococcus haemolyticus*.

REFERENCES

- Sasidharan N. Biodiversity documentation of Kerala, Part 6: Flowering Plants. Kerala Forest Research Institute, Peechi; 2004.
- Chethan J, Sampath Kumara KK, Niranjana SR, Prakash HS. Evaluation of antioxidant and antibacterial activities of methanolic flower extract of *Wedelia trilobata* (L.) Hitch. Afr J Biotechnol 2012; 11: 9829-9834. http://dx.doi.org/10.5897/AJB11.3729
- 3. Li D, Liang Z, Guo M, Zhou J, Yang X, Xu J. Study on the chemical composition and extraction technology optimization of

essential oil from *Wedelia trilobata* (L.) Hitchc. Afr J Biotechnol 2012; 11: 4513-4517.

- Neelam B, Nadpi GK, Titpawan N, Kholeeyoh J, Teerapol S. Evaluation of the wound healing potential of *Wedelia trilobata* (L.) leaves. J Ethnopharmacol 2012; 141: 817–824. http://dx.doi.org/ 10.1016/j.jep.2012.03.019
- Balekar N, Katkam NG, Nakpheng T, Jehtae K, Srichana T. Evaluation of the wound healing potential of *Wedelia trilobata* (L.) leaves. J Ethnopharmacol 2012; 141: 817-24. http://dx.doi.org/ 10.1016/j.jep.2012.03.019
- Balekar N, Nakpheng T, Katkam NG, Srichana T. Wound healing activity of ent-kaura-9(11), 16-dien-19-oic acid isolated from *Wedelia trilobata* (L.) leaves. Phytomedicine 2012; 19: 1178-84. http://dx.doi.org/10.1016/j.phymed.2012.07.014
- Taddei A, Rosas Romero AJ. Antimicrobial activity of *Wedelia* trilobata crude extracts. Phytomedicine 1999; 6: 133–134. http://dx.doi.org/10.1016/S0944-7113(99)80048-8
- Tambe VD, Nirmal SA, Jadhav RS, Ghogare PB, Bhalke RD, Girme AS, Bhambar RS. Anthelmintic activity of *Wedelia trilobata* leaves. Ind J Nat Prod 2006; 22: 27-29.
- Quang TT, Jean J, Akino J, Phi PNK, Ginette J. Wedelolides A and B: novel sesquiterpene δ-lactones, (9R)-eudesman-9,12-olides, from *Wedelia trilobata*. J Org Chem 2007; 72: 7102–7105. http://dx.doi.org/10.1021/j0070771m
- Nie C, Zeng R, Luo S, Li H, Hong M, Cheng L. Allopathic potentials of *Wedelia trilobata* L. on rice. Zuo wu xue bao 2004; 30: 942-946.
- 11. Raghavendra MP, Satish S, Raveesha KA. *In vitro* evaluation of anti-bacterial spectrum and phytochemical analysis of *Acacia nilotica*. J Agr Sci 2006; 2: 77-88.
- Bauer AW, Kirby MDK, Sherris JC, Turck M. Antibiotic susceptibility testing by standardized single disc diffusion method. Am J Clin Path 1996; 45: 493-496.
- 13. Barry AL. The Anti microbic Susceptibility Tests: Principles and Practices, Lea and Febiger Philadelphia; 1976.
- Ratimi VO, Laughon BE, Barlet JS, Mosadomi HA. Activities of Nigerian chewing sticks extract against *Bacterioides gingivalis* and *Bacterioides melaninogenicus*. Antimicrob Agents Chemother 1988; 32: 598-600. http://dx.doi.org/10.1128/AAC.32.4.598
- 15. Harborne JB. Phytochemical methods 1st ed. Chapman and Hall Ltd., London; 1973.
- Yue Zhong S. Recent Natural Products Based Drug Development: A Pharmaceutical Industry Perspective. J Nat Prod 1998; 61: 1053–1071. http://dx.doi.org/10.1021/np9800102
- 17. Doughari JH, El Mahmood AM, Jyoyina I. Antimicrobial activity of leaf extracts of *Senna obtusifolia* (L). Af J Pharm and Pharmacol 2008; 2: 7-13.
- Zhang L, Hua Z, Song Y, Feng C. Monoterpenoid indole alkaloids from *Alstonia rupestris* with cytotoxic, antibacterial and antifungal

activities. Fitoterapia 2014; 97: 142-147. http://dx.doi.org/ 10.1016/j.fitote.2014.05.018

- Masnon FF, Hassan NP, Ahmad F. Aporphine alkaloids of *Cinnamomum mollissimum* and their bioactivities. Nat Prod Commun 2014; 9: 31-32.
- Zhou XL, Xiao CJ, Wu LB, Huang B, Dong X, Jiang B. Five new terpenoids from the rhizomes of *Isodon adenantha*. J Asian Nat Prod Res 2014; 16: 555-564. http://dx.doi.org/10.1080/10286020. 2014.918110
- 21. Govindappa M, Naga Sravya S, Poojashri MN, Sadananda TS, Chandrappa CP. Antimicrobial antioxidant and *in vitro* antiinflammatory activity of ethanol extract and active phytochemical screening of *Wedelia trilobata* (L.) Hitchc. J Pharmacognosy Phytother 2011; 3: 43-51.

Cite this article as:

Rajalakshmi Shankar and Toji Thomas. Antibacterial activity of flower heads of Wedelia trilobata (L.) A. S. Hitche. J Biol Sci Opin 2014;2(6):409-412 <u>http://dx.doi.org/10.7897/2321-6328.02692</u>

Source of support: Nil; Conflict of interest: None Declared